

Two Cases of Vulvovaginitis Caused by *Shigella flexneri* and *Shigella sonnei*: a Case Report

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ABSTRACT

Vulvovaginitis caused by *Shigella* species (*Shigella* spp.) has rarely been reported. This paper describes two cases of prepubertal vulvovaginitis, presenting with a bloody and purulent vaginal discharge, separately caused by ampicillin-resistant *Shigella flexneri* and trimethoprim-sulfamethoxazole-resistant *Shigella sonnei*. Our conclusions are that *Shigella* spp. is the potential cause of vulvovaginitis in prepubertal girls in developing countries where these pathogens are endemic, and identification of the bacteria and making antibiotic susceptibility testing in these cases should not be overlooked.

Key Words: *Shigella flexneri*, *Shigella sonnei*, vulvovaginitis, prepubertal girls

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Introduction

Shigellosis or bacillary dysentery is a global human health problem with an estimated annual incidence of 164.7 million cases, of which 163.2 million occur in developing countries, and causes 1.1 million deaths (1). *Shigella* species are not part of the normal human flora, and are well-recognized agents of acute bacillary dysentery (2). Vulvovaginitis caused by *Shigella* spp. have been rarely reported (3-9). This may be partly due to the fact that diagnosis of *Shigella* vulvovaginitis may be difficult. Most of these cases are reported in children (3-9).

We describe here two cases of vulvovaginitis caused by *Shigella flexneri* and *Shigella sonnei*, and discuss some problems of the diagnosis and treatment of vulvovaginitis caused by *Shigella* spp.

Case Reports

Case 1

A nine-year-old girl was referred to our department for further evaluation of dyslipidemia and purulent vaginal discharge of unknown etiology. She had a history of a bloody-purulent vaginal discharge, and vulvovaginal itching for two months. There were no findings of any sexual abuse, history of serious illness and hospitalization, or the presence of any foreign body.

Vulvovaginal examination showed mucosal erythema with a yellowish purulent vaginal discharge mixed with blood. Gram stain smear of the vaginal discharge showed numerous polymorphonuclear leucocytes, gram-negative bacilli and predominant growth of *Escherichia coli*. No intracellular

gram-negative diplococci were seen at microscopic examination. The urine culture was negative. Based on the diagnosis of nonspecific vaginitis, the patient was advised on local hygienic measures. Since *E. coli* growth might have been due to contamination from the perianal skin, vaginal culture was repeated five days later and *S. flexneri* was identified. The third vaginal culture was again positive for *S. flexneri*. The stool cultures of the patient and other family members were negative for *S. flexneri*. The *S. flexneri* isolate was susceptible to ceftriaxone, ceftazidime, cefepime, ciprofloxacin, levofloxacin, chloramphenicol, trimethoprim-sulfamethoxazole, imipenem, meropenem and tigecycline and resistant to ampicillin and amoxicillin-clavulanic acid. The patient was treated with oral trimethoprim-sulfamethoxazole for 10 days, and follow-up vaginal cultures were found negative.

Case 2

An eight-year-old girl presented at our department with a one-year history of intermittent foul smelling, yellowish purulent vaginal discharge and severe vulvovaginal itching. The discharge had turned bloody during the last two months. Although treated elsewhere with unknown oral medication for one year, her symptoms had not improved. No other health problem, sexual abuse or trauma was reported.

The physical examination at admission showed a reddish vaginal mucosa and bloody-vaginal discharge. No vaginal foreign body was detected. *S. sonnei* resistant *in vitro* to trimethoprim-sulfamethoxazole but susceptible to ampicillin, amoxicillin-clavulanic acid, ceftriaxone, ceftazidime, cefepime, ciprofloxacin, levofloxacin, chloramphenicol, imipe-

nem, meropenem and tigecycline grew in the vaginal culture. The Gram-stained smear of the vaginal discharge showed numerous polymorphonuclear leucocytes but no gram-negative diplococci. The stool cultures of the patient and family members were negative for *Shigella* spp. and any other enteric pathogens. She was treated orally with amoxicillin-clavulanic acid for 10 days. The vaginal discharge was not resolved after 10 days of the treatment. On the third day after completion of antibiotic therapy, *S. sonnei* at 100 cfu per milliliter with the same susceptibility pattern was isolated from urine, but the repeated cultures from the vaginal discharge were negative. No further culture was performed since the patient made no further visits to our clinic for a month. When she returned after one month, vaginal culture had remained positive, and stool and urine cultures were negative for *S. sonnei*. The patient was prescribed oral cefixime for 10 days, and at follow-up examination, the vaginal culture was positive for *S. sonnei*. The patient was given a 14-day course of ciprofloxacin. The vaginal discharge resolved after the treatment with ciprofloxacin. Follow-up vaginal cultures were negative.

Discussion

Chronic vaginal discharge is the most frequent finding in *Shigella* vulvovaginitis and, as also noted with our cases, is resolved in longer than 10 days. The purulent-mucopurulent foul smelling discharge, varying from whitish to yellowish to greenish, does not differ from vulvovaginitis caused by the other microorganisms, however, the bloody discharge could be an indication for *Shigella* vulvovaginitis (5, 9). As shown in Table 1, eight out of 11 cases (73%), including ours, presented with bloody-vaginal discharge. Here no significant difference between *S. flexneri* and *S. sonnei* infection regarding the

frequency of bloody discharge has been detected, although bloody diarrhea due to *S. flexneri* has been reported more often compared to that of *S. sonnei* (9).

Laboratory diagnosis of *Shigella* vaginal infections is not easy. One of the possible reasons is that a request for the identification of *Shigella* is not common since many microbiology laboratories may not identify gram-negative rods recovered from vaginal cultures (4, 7). For instance to isolate and differentiate *Shigella* spp. in samples containing bacterial flora, it is advised that clinical sample should be inoculated onto Salmonella Shigella (SS) agar, eosin-methylene blue (EMB) and selenite broth (SB) and incubated for longer times (10). None of these growth media are mentioned among the media suggested for routine culturing of vaginal samples. When it has been done, the suggested media for vaginal specimens of children are 5% sheep blood agar and chocolate agar for *Streptococcus pyogenes*, *Streptococcus pneumoniae* and *Haemophilus influenzae*, Sabouraud medium for *Candida* spp. and selective gonococcal medium for *Neisseria gonorrhoeae*. Because, *Shigella* spp. and other gram-negative bacilli are not considered as possible vulvovaginitis pathogens in prepubertal girls, (11) and gram-negative rods (such as *Proteus* species, *Pseudomonas* species) known to occur in healthy prepubertal girls without causing any illness (12). Additionally, Gram-negative rods of fecal origin can also be found as contaminants. As a result, *Shigella* spp. is easily overlooked in routine vaginal cultures if no pure or prominent growth was detected. Another reason for missing *Shigella* in laboratory diagnosis is inaccurate specimen collection and transportation. *Shigella* spp. are fragile organisms and remain viable for a limited time outside the human body. Therefore, considerable care must be taken in the collection and transportation of the clinical specimens, and informing the laboratory about the possible causative

Table 1. Reported cases (Including the Present Cases) of *Shigella* vulvovaginitis. At the far left column, the number of the cases; at the top line, the features of the cases have been shown.

Case	Year	Ref.	Age	<i>Shigella</i> species	Bloody discharge	Associated symptoms	Antibiotic resistance pattern	Ineffective treatment	Effective treatment
1	1950	(3)	7	<i>S. flexneri</i>	No	No	ND	No	Sulphaguanidine orally
2	1975	(4)	6	<i>S. flexneri</i>	Yes	No	ND	No	Ampicillin orally
3	1975	(4)	4	<i>S. flexneri</i>	Yes	Urinary frequency and dysuria	ND	No	Ampicillin orally
4	1975	(4)	4	<i>S. flexneri</i>	Yes	Dysuria	ND	No	Ampicillin orally
5	1975	(4)	6	<i>S. flexneri</i>	Yes	No	ND	Nitrofurantoin, ampicillin orally	Estrogen/sulfasoxazole intravaginally
6	1979	(5)	4	<i>S. sonnei</i>	No	No	Ampicillin	Ampicillin orally	Estrogen/sulfasoxazole intravaginally
7	2002	(6)	7	<i>S. flexneri</i>	Yes	Dysuria	Ampicillin, SXT	SXT, AMC, cefixime orally	Ciprofloxacin orally
8	2002	(7)	5	<i>S. flexneri</i>	Yes	No	No resistance	No	SXT orally
9	2006	(8)	5	<i>S. flexneri</i>	No	Dysuria	Ampicillin, SXT	No	Cefixime orally
10*	2009		9	<i>S. flexneri</i>	Yes	No	Ampicillin, AMC	No	SXT orally
11*	2009		8	<i>S. sonnei</i>	Yes	No	SXT	AMC, cefixime orally	Ciprofloxacin orally

*The last two cases are described in the present paper, ND=Not described, SXT=Trimethoprim-sulfamethoxazole, AMC=Amoxicillin-clavulanic acid

agents (13). For a fast and accurate diagnosis, the microbiology laboratory should be certainly informed by clinicians if a *Shigella* infection is considered or if vaginal discharge with blood is present in the patient. The presence of leucocytes in vaginal secretions may be an indicator for clinicians.

Because of the infrequency of the disease, and the limitations in clinical experience, the optimal therapy for *Shigella* vulvovaginitis remains uncertain (7, 8). Systemic antibiotic therapy is generally recommended as being more effective than topical antibiotic therapy (4, 6, 8). Although the recommended standard course of therapy for shigellosis is oral ampicillin or trimethoprim-sulfamethoxazole therapy, it should be noted that, while initially susceptible, most *Shigella* isolates are currently resistant to these antibiotics. These are inappropriate for empiric therapy unless local microbiological data suggest susceptibility (1, 6, 8).

In the treatment of shigellosis, intracellular concentrations of antibiotics are probably the crucial determinants of success, since *Shigella* spp. multiply intracellularly. Fluoroquinolones such as ciprofloxacin, and macrolides such as azithromycin, may be particularly useful for the treatment of shigellosis, because, they reach high intracellular levels (13, 14). On the other hand, the use of fluoroquinolone in children has been limited because of the concerns about its toxicity. However, there is growing evidence of their safety for the treatment of shigellosis in children (1, 12). Azithromycin is approved for use in children (13, 14). The significant disadvantage of the azithromycin is the lack of Clinical Laboratory Standards Institute-defined breakpoints for it for the *Enterobacteriaceae*, including *Shigella* spp. (15). Susceptibility of the *Shigella* isolates to azithromycin is not routinely tested in most countries which use Clinical Laboratory Standards Institute documents for antibiotic susceptibility tests, as in Turkey.

In our first case, the *S. flexneri* isolate was susceptible to a variety of agents including trimethoprim-sulfamethoxazole, and the infection was successfully treated with oral trimethoprim-sulfamethoxazole. In our second case, however, the *S. sonnei* isolate was susceptible *in vitro* to amoxicillin/clavulanic acid and cefixime, but the treatments with these antibiotics were not successful. Failure with amoxicillin/clavulanic acid and cefixime was also reported by Baiulescu et al. (6) in a child with *Shigella* vulvovaginitis, in which the organism was susceptible *in vitro* to amoxicillin/clavulanic acid and cefixime, but the patient exhibited clinical and microbiologic failure until ciprofloxacin therapy was instituted. Our case and the case reported by Baiulescu et al. (6) differ only in respect to the agents isolated, *S. sonnei* and *S. flexneri*, respectively. The failures in treatment with amoxicillin/clavulanic acid and cefixime suggest a possible difference in the form of infection rather than *Shigella* species. It is likely that involvement of gynecological tissue complicates the success of the management.

In conclusion, our experience strongly suggests that identification and antibiotic susceptibility testing for *Shigella* spp., as a potential cause of vulvovaginitis in prepubertal girls presenting with chronic vaginal discharge, especially in developing countries where these pathogens are endemic, should not be overlooked. Additionally, our current understanding suggests that fluoroquinolone antibiotics such as ciprofloxacin might not be the first but nevertheless the best choice against vulvovaginitis for the patients not responding to the regular treatment, and there is not always a good correlation between *in vivo* clinical response and *in vitro* antibiotic susceptibility test results.

Conflict of Interest

No conflict of interest was declared by the authors.

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